Tetrahedron Letters No.11, pp. 1211-1218, 1966. Pergamon Press Ltd. Printed in Great Britain.

## THE CONSTITUTION OF COCHLIOBOLIN

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(Received 17 December 1965; in revised form 21 January 1966)

We propose the formula <u>1</u> for the cochliobolin,  $C_{25}H_{36}O_4$  (MS 400), a metabolic product of Helminthosporium orizae:



<u>1</u> has m.p.  $181^{\circ}$ ;  $(a)_D^{20}$  301; UV 236, 10100; IR (CHCl<sub>3</sub>) 3450, 2730, 1742, 1673, 1635; its NMR spectrum shows signals at 0.83 s C11-CH<sub>3</sub>, 1.12 d(7) (DR 2.28 s), C15-CH<sub>3</sub>, 1.37 s C3-CH<sub>3</sub>, 1.78 broad s (DR 5.18 s) two C19-CH<sub>3</sub>, 2.65 AB q(20) C4-H<sub>2</sub>, 3.25 d(10) (DR 2.28 s) OH C6-H, 4.45 m (DR 1.75 d, 5.18 t) C17-H, 5.18 broad d(7) (DR 1.78 d, 4.45 s) C18-H, 7.21 t (DR 2.22 s) C8-H, 9.23 s C21-H. <u>1</u> yields a monoepoxide and an anhydrobis-2,4-dinitrophenylhydrazone m.p. 252°; in fact <u>1</u> is dehydrated in acid and alkaline medium to 3-anhydrocochlio-

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a) MS indicates the determination of the molecular weight by mass spectrometry; (a) were determined in chloroform; UV spectra were run in methanol ( $\lambda_{max}$  in me, intensities as  $\varepsilon$ ); IR spectra in nujcl and  $\gamma_{max}$  in cm<sup>-1</sup>; NMR spectra were determined in CDCl, (at 60 Mc, TMS as internal reference), chemical shifts as  $\delta$ =ppm (J in cps); DR signifies a double resonance spectrum at the indicated frequency.

bolin, C<sub>25</sub>H<sub>34</sub>O<sub>3</sub>; MS 382; m.p. 135°; (a)<sub>D</sub><sup>20</sup> 164; UV 232, 20900; IR 2700, 1695, 1675, 1640, 1620; NMR spectrum of the 3-anhydro derivative shows disappearance of the signals at 1.37, 2.65, 3.25, present in 1, while new signals at 2.09 broad a C3-CH, 3.47 d(5) C6-H, 6.11 s C4-H are observed. With LiAlH<sub>4</sub> <u>1</u> yields two stereoisometric triols  $C_{25}H_{40}O_4$ , <u>2</u> and 3: 2 (MS 404); m.p. 158-60° (uncrystallizable dibenzoate) and 3 (MS 404); m.p. 176-78° (dibenzoate m.p. 148-51°). 2 yields 1 with CrO, in pyridine and 4 with  $MnO_2$ . 4  $C_{25}H_{38}O_4$ shows UV 240, 8000; IR 3500, 3350, 1675, 1630. 3 with Cro, in pyridine or with MnO<sub>2</sub> yields 5 C<sub>25</sub>H<sub>36</sub>O<sub>4</sub>; MS 400; m.p. 182-4°; UV 227, 10050; IR (CHCl<sub>3</sub>) 3700, 3500, 1750, 1685; NMR spectrum of 5 shows disappearance of the signals at 1.37, 1.78, 2.65, 3.25, 7.21, 9.23 present in 1 while new signals at 1.25 s C3-CH<sub>2</sub>, 1.5 OH, 1.70 broad s two C19-CH<sub>2</sub>, 3.56 m C6-H, 4.9 m C5-H, 7.0 m C8-H are observed. 5 yields 3 with LiAlH<sub>4</sub>. On hydrogenation 5 yields 6, C25H4004; MS 404; m.p. 126°; IR 3510, 1740; MMR spectrum of 6 shows disappearance of the signals at 1.70, 3.56, 4.45, 4.9, 5.18, 7.0 present in 5 while new signals at 0.89 d(6) two C19-CH<sub>2</sub>, 3.8 m C17-H, 5.07 m C5-H are observed. These results are in agreement with the following scheme:



On hydrogenation 1 yields 7,  $0_{25}H_{40}O_4$ ; MS 404; m.p. 156-7°; IR (CHCl<sub>2</sub>) 3340, 3110, 1740, 1663; NMR spectrum of <u>7</u> shows disappearance of the signals at 1.78, 2.65, 3.25, 4.45, 5.18, 7.21, 9.23 present in 1, while new signals at 0.91 d(7) two C19-CH<sub>2</sub>, 2.52 B C4-H<sub>2</sub>, 2.9 d(10) C6-H, 3.76 m C17-H, 4.8 broad s OH, 6.3 m C21-H, 8.63 m OH are observed. 7 yields a mono-4-nitrobenzoate C32H43NO7; m.p. 132-5° which shows two signals at 3.38 d(7) and 7.18 broad s in its NMR spectrum (respectively at 2.9 and 6.3 in  $\underline{7}$ ). With acids such a derivative yields an isomer m.p. 185-7° which shows two signals at 3.6 d(12) and 7.42 s. Both the 4-nitrobenzoates yield the 3-anhydroderivatives with acids. With LiAlH, 7 yields two isomeric hemiacetals, which yield 6 with Cr0, in pyridine. With 2,3-dichloro-5,6-dicyanobenzoquinone 7 yields 8,  $C_{2c}H_{20}O_4$ ; the UV spectrum of that is identical to that of 1 and the IR spectrum differs only in the fingerprint region; NMR spectrum of 7 shows disappearance of the signals at 1.78. 4.45, 5.18 present in 1 while new signals at 0.92 d(6) two C19-CH, and 3.74 m C17-H are observed. Owing to these reasons we suggest the tentative formula 7. In this particular case the aldehyde group is stabilized in enol form by steric influences; the spect oscopic properties of 7 suggest that the C3-OH is involved in this stabilization through a hydrogen bond.



With 0, on Pd-C 7 yields a peroxide having the tentative for-

mula 9  $C_{25}H_{40}O_6$  (MS: m/e = 374 = P - 62); m.p. 115°; IR 3500, 3450; NMR spectrum of 9 shows disappearance of the signals at 1.37, 2.52, 2.9, 4.8, 6.3, 8.63 present in 1 and 7 while new signals at 1.24 s C3-CH<sub>3</sub>, 3.27 d(5) C6-H, 4.02 d(13.4) C21-OH, 5.06 d(13.4) C21-H, 5.88 OH are observed.



In fact with  $SO_2$  (with neither  $SO_3^-$  nor  $HSO_3^-$ ) 9 yields 10 quickly; by-products were observed, but no  $H_2SO_4$  was formed. 10 C<sub>25</sub>H<sub>38</sub>O<sub>5</sub>, has m.p. 120-2°; UV 290, 9600 (in alkali 313, 15500); IR 1720, 1650, 1610, 1190; NER spectrum of 10 shows signals at 0.83 s C11-CH3, 0.98 d(7) C15-CH3 and two C19-CH3, 1.62 B C3-CH<sub>3</sub>, 2.8 s C4-H<sub>2</sub>, 3.04 m C2-H, 3.86 m C17-H, 8.05 s C21-H. By saponification, 10 yields formic acid and 11 C<sub>24</sub>H<sub>38</sub>C<sub>4</sub>; MS 390; m.p. 85-96°; UV 290, 9600 (in alkali 313, 17700); IR (CHCl<sub>3</sub>) 3620, 3430, 1750, 1660, 1610; NMR spectrum of 11 shows disappearance of the signals at 1.62, 2.8, 8.05 present in 10 while signals at 1.38 s C3-CH, 2.44 s C4-H<sub>2</sub> are observed. With hydrazine <u>11</u> yields a pyrazole C<sub>24</sub>H<sub>38</sub>N<sub>2</sub>O<sub>2</sub>; MS 386; m.p. 109-112°; UV 226, 6730 (in acidic medium 233, 3500). By deformylation of 10 or dehydration of <u>11</u> we obtain <u>12</u>  $C_{24}H_{36}O_3$ ; m.p. 102-4°; UV 245, 8140 - 310, 5500 (in alkaline medium 335, 9350); NMR spectrum of <u>12</u> shows disappearance of the signals at 1.62, 2.8, 3.04, 8.05 present in 10 while new signals at 2.09 s C3-CH<sub>3</sub>, 3.14 m

C2-H, 6.02 s C4-H are observed. <u>11</u> is also obtained from <u>7</u> with  $0_2$  and FeCl<sub>3</sub> and from <u>9</u> with FeCl<sub>3</sub>. Owing to these results we can explain this reaction as follows:



With one mole of  $HIO_4$  9 yields quantitatively formic acid and a monocarboxylic acid 13  $C_{24}H_{38}O_5$ ; m.p. 162-5° which yields a methyl ester 14  $C_{25}H_{40}O_5$ ; MS 420; b.p.  $170^{\circ}/10^{-4}$  Torr; IR 1760, 1740; NMR spectrum of 14 shows signals at 0.8 s C11-CH<sub>3</sub>, 0.88 d(6) two C19-CH<sub>3</sub>, 0.99 d(7) C15-CH<sub>3</sub>, 1.37 s C3-CH<sub>3</sub>, 2.7 s C4-H<sub>2</sub>, 2.8 m C2-H and C6-H, 3.68 s CH<sub>3</sub> ester, 3.87 m C17-H. Its mass spectrum shows a peak at m/e = 347 = P - CH<sub>2</sub>COOCH<sub>3</sub>. From the above data we can explain the reaction with HIO<sub>4</sub> as shown in the scheme at the next page. In alkaline medium 9 yields the acid 15  $C_{24}H_{38}O_5$ ; m.p. 214-7°; physical properties of 15 (and of its methyl ester 16  $C_{25}H_{40}O_5$ b.p. 170°/10<sup>-4</sup>Torr) are very similar to the 13 and 14 ones.



<u>15</u> is also obtained from <u>11</u> with  $H_2O_2$  in acetic acid. We explain the reaction in this manner:

With sodium methoxide in methanol <u>14</u> and <u>16</u> yield a mixture of two dicarboxylic mono- $a,\beta$ -insaturated acids C<sub>24</sub>H<sub>38</sub>O<sub>5</sub>; m.p. 80-90°; UV 225, 11700.

On hydrogenation <u>3</u> yields <u>17</u>  $C_{25}H_{44}O_3$ ; MS 392; m.p. 160-3°; IR 3570, 3470, 3430; NMR spectrum shows signals at 0.8 d(6) C15-CH<sub>3</sub>, 0.87 s C11-CH<sub>3</sub>, 0.88 d(6) two C19-CH<sub>3</sub>, 1.32 s C3-CH<sub>3</sub>, 1.55 30H, 2.05 s C7-CH<sub>3</sub>, 3.18 m C6-H, 4.45 m C5-H, 6.65 m C8-H. Its mass spectrum shows the main peak m/e = 225 \* P - $-C_8H_{17} - 3H_2O$ , which suggests the presence of a saturated aliphatic chain  $C_8H_{17}$ . All the other derivatives of <u>1</u> we investigated by mass spectrometry have no significant peaks at m/e = P -  $C_8H_{17} - nH_2O$ , but the most abundant peak is at  $m/e = 165 = C_{11}H_{17}^{0}$  for the derivatives having an isopropylidene group and  $m/e = 167 = C_{11}H_{19}^{0}$ , <u>18</u>, for ones having an isopropylic group.

Owing to these results we suggest that on hydrogenation of  $\underline{3}$  the ether bridge between C14 and C17 was cleaved the  $\Delta_{18}$  hydrogenated and the C21-H<sub>2</sub>OH group hydrogenolyzed to form a methyl group.



With perbenzoic acid, acid hydrolysis and  $\text{HIO}_4 \ \underline{1}$  yields acetone; with  $\text{HNO}_3 \ \underline{1}$  gives  $\underline{19}$  and all its nor-derivatives;  $\underline{19}$ can be also obtained from  $\underline{7}$  with the lactone  $\underline{20}$ ;  $\underline{17}$  yields  $\underline{19}$ , all its nor-derivatives and  $\underline{21}$ .  $\underline{19}$ ,  $\underline{20}$ ,  $\underline{21}$ , show IR, NMR MS spectra identical to synthetized products of unequivocal structure. With  $\text{HNO}_3 \ \underline{7}$  does not yield acids having a long)r chain than  $\underline{19}$ ; this result suggests that in the original structure the quaternary methyl group, appearing in  $\underline{19}$ , must be separated from the  $\Delta_{\ \underline{7}}$  by a trisubstituted carbon atom. Therefore, for cochliobolin the constitutions  $\underline{1}$  or  $\underline{22}$  are possible only.



We propose the formula 1 for cochliobolin since such an ion

as <u>18</u>, occuring in its mass spectrum, cannot be easily explained according to <u>22</u>. Recently Nozoe et al.<sup>2</sup> deduced for ophiobolin the structure <u>1</u>, including the absolute configuration by X-ray crystallographic analysis of its bromomethoxy derivative <u>23</u>. We have observed that physical constants of cochliobolin and of its bromomethoxy derivative are very similar with the reported constants of ophiobolin and of its bromomethoxy derivative<sup>2</sup> and we agree with Nozoe et al. in suspecting the identity of cochliobolin with ophiobolin.

<u>Acknowledgement</u>. - We are very grateful to drs. G. Nencini and T. Salvatori of Laboratori Riuniti SNAM for their decisive interpretations of mass spectra.

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